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REVIEW ARTICLE

Microbes and the gut-brain axis

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Abstract

Background The 'gut-brain' or 'brain-gut axis', depending on whether we emphasize bottom-up or top-bottom pathways, is a bi-directional communication system, comprised of neural pathways, such as the enteric nervous system (ENS), vagus, sympathetic and spinal nerves, and humoral pathways, which include cytokines, hormones, and neuropeptides as signaling molecules. Recent evidence, mainly arising from animal models, supports a role of microbes as signaling components in the gut-brain axis. Aims The purpose of this review is to summarize our current knowledge regarding the role of microbes, including commensals, probiotics and gastrointestinal pathogens, in bottom-up pathways of communication in the gut-brain axis. Although this has clear implications for psychiatric co-morbidity in functional and inflammatory conditions of the gut, the focus of this review will be to discuss the current evidence for a role of bacteria (commensals, probiotics, and pathogens) as key modulators of gut-brain communication. Results € Conclusions The strongest evidence for a role of microbes as signaling components in the gut-brain axis currently arises from animal studies and indicate that mechanisms of communication are likely to be multiple. There is need for the concepts generated in animal models to be translated to the human in the future.

INTRODUCTION

Clinicians and researchers have long recognized the link between gastrointestinal function and the central

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Elena F. Verdu, 1280 main Street West, HSc 3N8, Hamilton, ON, L8S 4K1, Canada. Tel: 905 525-9140x20051; fax: 905 522-3454; e-mail: verdue@mcmaster.ca *Received*: 9 January 2012 *Accepted for publication*: 13 Febraury 2012 nervous system (CNS). Although the original description of a gut-brain axis related to the modulation of cholecystokinin secretion by bombesin,¹ the concept has since then been extended to describe any interaction between the gastrointestinal tract and the CNS. Recently, results in animal models have generated great interest into the role of intestinal microbes as key players in gut-brain communication (Fig. 1). The neural aspects and the role of centrally-driven pathways in gut-brain axis communication have recently been reviewed in detail by Mayer *et al.*² and O'Mahony *et al.*³

Intestinal microbiota and gut homeostasis

The intestinal microbiota involves a wide diversity of microbial species⁴ and can be considered a postnatal acquired organ that performs different functions for the host. Intestinal microbes have developed a mutualistic relationship with its host and play a crucial role in the development of innate and adaptive immune responses,^{5,6} influence physiological systems throughout life by modulating gut motility, intestinal barrier homeostasis,^{7,8} absorption of nutrients and the distribution of somatic and visceral fat.^{9,10}

The intestinal microbiota consists of a community of bacteria that colonize the gastrointestinal tract after birth and persist throughout adult life, and 'transient' bacteria, such as probiotic bacteria, which are temporarily acquired during ingestion of certain foods. The composition of the intestinal microbiota is established during the first few years of life and is likely shaped by multiple factors including maternal vertical transmission, genetic make up of the individual, diet, medications such as antibiotics, gastrointestinal infections and stress^{11–15} (Fig. 2). Until recently composition of this microbial community was considered unique for each individual and relatively stable over time.^{16,17} However, using deep sequencing of stool samples from



Figure 1 The intestinal microbiome: a complex and dynamic ecosystem that establishes bi-directional interactions with the host.

In addition to the known effects of intestinal microbiota and specific probiotics on mucosal and epithelial barrier function, there is experimental evidence to support the effects of microbes on muscle function.^{1–5} and ENS function.^{6–9} The information is relayed to the CNS via neural^{10–12} or humoral pathways.⁹

several hundred individuals, the European MetaHit consortium study has shown that human microbiota profiles can be grouped in three major bacterial enterotypes dominated by *Bacteroides*, *Prevotella* and *Ruminococcus*, respectively.¹⁸ Existence of distinct enterotypes strongly associated with longterm diets has been confirmed by Wu *et al.*, linking protein and animal fat with *Bacteroides* and consumption of carbohydrates with Prevotella.¹⁹ This indicates that despite existence of large number of bacterial strains in the human intestine there are only a limited number of well-balanced host-microbial symbiotic states that might respond differently to diet and drug intake.

Although the intestinal microbiota is considered relatively stable, some studies have shown a marked

Figure 2 The gut-brain axis. Pathways of communication and probiotic targets.

variation in the complexity and stability of Bifidobacteria and Lactobacillus populations over a 12-month period²⁰ and significant alterations in microbiota composition due to environmental factors.^{21,22} Treatment with antibiotics is known to affect the intestinal microbiota and these changes may be long lasting. A study investigating the effect of a short course of amoxicillin showed that the microbial ecology of the intestinal tract was severely altered for up to 6 months after the end of antibiotic administration.²³ Similarly, 1-week course of clindamycin caused changes in the Bacteroides community, and this persisted for up to 2 years.^{24,25} Diet is also likely to have a profound effect on gut bacterial composition. Different bacterial phenotypic profiles were found in subjects consuming either a Western style or an agrarian diet on a chronic basis.²⁶ Although it would be logical to expect that acute changes in diet would alter the gut microbiota, recent short-term experiments in healthy volunteers using either high fat of high carbohydrate diets have not shown any marked changes in bacterial enterotypes.¹⁹ It is thus possible that a stable microbiota is a hallmark of health, as unstable microbiota profiles have been reported in patients with irritable bowel syndrome and inflammatory bowel disease.^{27–30}

Intestinal microbiota and the gut brain axis

Indirect evidence for an effect of microbiota on CNS: clinical studies The concept that gut bacteria are a driving force for immune maturation and gut function in the host is well accepted. The notion that bacteria could also influence brain function and behavior is seemingly implausible, but clinicians use on a routine basis laxatives and oral antibiotics to treat patients with altered mental status due to hepatic encephalopathy.³¹ Several clinical studies have also described altered composition of gut microbiota in patients with autism³² and suggested at least short-term beneficial effect of antibiotic treatment,^{33,34} although there is no randomized clinical trial available to date. There are also multiple reports of patients developing psychosis after administration of different antibiotics.³⁵ To our knowledge, there have been no studies that characterize the gut microbiota associated with depression or anxiety, but earlier studies demonstrated that depression in females is associated with increased fermentation of carbohydrates, indirectly implicating changes in the composition or metabolic activity of the gut microbiota.36,37

Direct evidence for an effect of commensals and pathogens on CNS: animal studies At this point, the brunt of evidence linking microbes with behavior and brain biochemistry comes from animal studies. Pivotal experiments performed by Lyte *et al.* have shown that mice display altered, anxiety-like behavior during the early phase of acute infection with Campylobacter jejuni.38 This abnormal behavior occurred within several hours after introduction of the intestinal pathogen into the GI tract, before any significant immune response was mounted suggesting that this was not a consequence of cytokine-induced sickness behavior. Subsequent studies showed that presence of C. jejuni or Citrobacter rodentium triggers activity of vagal ascending pathways, such as nucleus tractus solitarius (NTS) and the lateral parabrachial nucleus,³⁹⁻⁴¹ and a specific activation pattern in multiple brain regions previously implicated in anxiety-like behavior.42 This

clearly illustrates that that the neural system can detect an acute change in the gut and selectively identifies the presence of a pathogen in the gut lumen.

Studies using chronic H. pylori infection in mice have shown that this pathogen alters gastric physiology, namely delayed gastric emptying and visceral sensitivity, with up-regulation of SP and CGRPcontaining nerves in the stomach and the spinal cord.^{43,44} Furthermore, chronic *H. pylori* infection leads to abnormal feeding behavior, characterized by frequent feeding bouts but with less food consumed per feeding bout than controls, which is reminiscent of early satiety observed in patients with functional dyspepsia.⁴⁴ The abnormal feeding pattern was accompanied by down-regulation of regulatory peptide Proopio-melanocortin (POMC) in the arcuate nucleus and up-regulation of the pro-inflammatory cytokine TNF-α in the median eminence (ME) of the hypothalamus. The ME is a part of the circumventricular organ, area of the brain where blood-brain barrier is relatively leaky enabling metabolites/molecules from the systemic circulation to enter the CNS. Interestingly, altered behavior and biochemical abnormalities persisted for at least two months postbacterial eradication suggesting that changes induced by chronic infection in the CNS may be long lasting or permanent.

To establish a link between commensal bacteria and the CNS, several experimental approaches can be undertaken. One is to compare germ-free with animals colonized with specific pathogen flora (SPF). Sudo et al. demonstrated an abnormal HPA axis with elevated ACTH and corticosterone levels in response to restraint stress in germ-free mice, which normalized after colonization with commensal bacteria.45 Furthermore, germ-free mice had lower brain derived neurotrophic factor (BDNF) levels in the cortex and hippocampus. Several recent studies have compared behavior and brain biochemistry in germ-free and SPF mice. Overall, using standard behavioral tests, such as elevated plus maze, open field and light/dark preference tests, germ-free mice displayed higher exploratory and lower anxiety-like behavior than SPF mice.46,47 Heijtz et al. showed that compared to germ-free mice, SPF mice had higher central expression of neurotrophins, such as nerve growth factor (NGF) and BDNF.⁴⁶ Furthermore, there was differential expression of multiple genes involved in the secondary messenger pathways and synaptic long-term potentiation in the hippocampus, frontal cortex and striatum. Similarly, Neufeld et al. demonstrated increased expression of NMDA receptor subunit NR2B in the central amygdala and serotonin receptor 1A (5-HT 1A) expression in the hippocampus in SPF mice compared to germ-free

mice.⁴⁷ The pronounced differences between germ-free mice and mice colonized with complex microbiota may relate to the ability of gut bacteria to affect multiple aspects of host metabolism, immunity and physiology. Colonization with a single commensal bacterium, *B. thetaiotaomicron* was shown to change expression of a vast array of genes in the intestine encoding for metabolism, intestinal permeability, angiogenesis but also for glutamate uptake, GABA production and neurotransmitter release.⁴⁸

A different approach to investigate the role of microbiota in gut-brain axis is to perturb a previously 'stable' microbiota in adult healthy mice by oral administration of non-absorbable antimicrobials. Combination of neomycin, bacitracin and pimaricin induced changes in colonic microbiota composition (gut dysbiosis) in SPF mice, with a marked increase in Firmicutes, mainly Lactobacilli spp, and decrease in γ -proteobacteria. This was accompanied by an increase in mouse exploratory behavior and altered BDNF levels in hippocampus and amygdala⁴⁹ (Fig. 3A). The same antimicrobial treatment failed to induce behavior abnormalities in germ-free conditions or in mice treated with antimicrobials intraperitoneally. The antimicrobial regime employed in this study did not induce measurable changes in gut inflammation or change levels of intestinal serotonin (5-HT), noradrenalin (NA) or dopamine. Interestingly, studies using subdiaphragnatic vagotomy or chemical sympathectomy before antimicrobials suggest that vagal and sympathetic pathways are not involved in gut-brain communication in this experimentally-induced dysbiosis model of altered behavior.

Behavior has a genetic component, and it is known that mouse strains differ in their behavioral phenotype.^{50,51} There is also a difference in microbiota composition among mouse strains, and the 'SPF' status does not indicate uniformity of the microbiota, only the fact that mice have been screened for the most common murine pathogens. BALB/c and NIH Swiss mice stand on opposite ends of the behavior phenotype: BALB/c mice are timid and less exploratory while NIH Swiss mice display a high exploratory drive. The BALB/c and NIH Swiss mice were reared under germ-free conditions and then colonized with SPF microbiota from either NIH Swiss or BALB/c mice. Germ-free mice colonized with microbiota from the same strain exhibited similar behavior as the SPF mice. However, mice colonized with microbiota from the other strain, exhibited a behavior profile similar to the donor⁴⁹ (Fig. 3B). This was not accompanied by any measurable change in systemic or gut immune activation or levels of intestinal 5-HT, NA or dopamine. A change in central neurotrophins was observed one week postcolonization. We can therefore speculate that host behavioral phenotype is also influenced by microbial factors.

The effect of microbiota may extend into memory and cognition, as recently suggested in a study comparing germ-free and SPF mice.⁵² An Earlier study by Li *et al.* examined the effects of long-term dietary manipulation on memory.⁵³ Mice assigned to chow with high content of ground beef for 3 months displayed higher diversity of gut microbiota compared to mice on standard rodent chow. This was associated with improved working and reference memory, as well



Figure 3 Effects of commensal bacteria on behavior. (A) Exploratory behavior of adult BALB/c mice increased transiently in parallel with gut dyshiosis induced by oral antimicrobial (ATM) treatment. This was accompanied by higher brain derived neurotrophic factor (BDNF) levels in the hippocampus. (B) Colonization of germ-free NIH Swiss and BALB/c mice with either NIH Swiss or BALB/c microbiota promoted or inhibited their exploratory behavior, respectively. Conventional, specific pathogen free (SPF) mice of both strains were used as controls. Adapted from Bercik et al., Gastroenterology, 2011 (reproduced with permission from Gastroenterology).

as slower speed in seeking food and anxiety-like behavior in mice.⁵³

Effects of probiotic bacteria on CNS and enteric nervous system function Psychiatric co-morbidities, such as anxiety and depression are common in patients with chronic bowel disorders, including IBS and inflammatory bowel disease.54-58 Both of these disorders are also associated with abnormal intestinal microbiota profiles.²⁷⁻³⁰ In this respect, chronic infection with a non-invasive parasite and mild chemically-induced colitis were shown to be associated with anxiety/depression-like behavior and decreased levels of hippocampal BDNF expression^{59,60} (Fig. 4). Interestingly, both abnormalities were normalized by the treatment with the probiotic B. longum NC3001 but not with L. rhamnosus NCC4007. Bifidobacterium longum did not improve gut inflammation or circulating cytokines, however, its anxiolytic effect was absent in mice with previous vagotomy, suggesting that its action was neurally mediated. This was further confirmed by ex vivo studies, in which electroresponsiveness of enteric neurons was assessed after perfusion with B. longum supernatant. Compared to controls, B. longum treated neurons fired less action potentials in response to supra-threshold depolarizing current.⁶⁰ The results suggest these signals may be initiated at the levels of

the enteric nervous system (ENS). The beneficial effect of probiotic bacteria may extend to healthy individuals. A study by Desbonnet et al.⁶¹ showed that administration of Bifidobacterium infantis to healthy Sprague-Dawley rats reduced concentrations of serotonin and dopamine metabolites in the frontal and the amygdaloid cortex, respectively. The authors suggested that this bacterium might have an anxiolytic potential, although no difference in behavior was found in that study. Subsequent experiments with the same bacterium using maternal separation model demonstrated beneficial effect on altered behavior together with normalization of noradrenaline concentrations in the brainstem.⁶² Bravo et al. have recently demonstrated that administration of the probiotic L. rhamnosus JB1 promoted exploratory behavior and attenuated despair-like behavior, as assessed by elevated plus maze and forced swim test, respectively, in healthy BALB/c mice. This was accompanied by a region-dependent alterations in GABA(B1b) and GABA(A α 2) mRNA in the brain,⁶³ which was vagally dependent, as subdiaphragmatic vagotomy abolished both changes in brain biochemistry and behavior. Thus, animal studies support the notion that commensal bacteria and specific probiotics can influence brain chemistry and the function of the CNS, perhaps by modulating the ENS. This has implications for pain perception. Visceral pain perception is regu-



Figure 4 Effects of single probiotic on central brain derived neurotrophic factor (BDNF) and anxiety-like behavior. (A) Representative micrographs of BDNF mRNA expression in the hippocampus using in situ hybridization. BDNF was normalized in mice with chronic T. muris colitis treated for 2 weeks with Bifidobacterium longum (B). (C) Anxiety-like behavior normalized in mice with T. muris colitis after treatment with B. longum, but not with Lactobacillus rhamnosus. Adapted from Bercik *et al.*, Gastroenterology, 2011 (reproduced with permission from Gastroenterology). lated by complex mechanisms, including peripheral sensitization of sensory nerves as well as central processing, which is modulated by concurrent anxiety and depression. Both of these processes could be affected by intestinal microbiota. There are multiple clinical trials that have reported improvement of pain after treatment with different probiotic bacteria (Table 1) but the mechanisms of action remain unknown. In one clinical trial, treatment with *Bifidobacterium infantis* normalized the increased ratio of serum pro-inflammatory cytokines suggesting an anti-inflammatory effect may mediate improvement in pain perception in IBS patients.⁶⁴

Animal models have provided evidence on possible neural and metabolic pathways affected by commensals and probiotics. The first report on the role of bacteria on

 Table 1 Randomized control trials showing beneficial effect of probiotics on abdominal pain

Reference	Probiotic treatment
Niedzielin <i>et al.</i> , Eur J Gastroenterol Hepatol 2001	Lactobacillus plantarum 299V
O'Mahony <i>et al.</i> , Gastroenterology 2005	Bifidobacterium infantis 35624
Bausserman et al., J Pediatr 2005	Lactobacillus GG
Whorwell et al., J Gastroenterol 2006	6Bifidobacterium infantis 35624
Gawronska <i>et al.</i> , Alim Pharmacol Ther 2007	Lactobacillus GG
Sinn et al., Dig Dis Sci 2008	Lactobacillus acidophilus SDC 2012
Drouault-Holowacz et al.,	B. longum LA101,
Gastroenterol Clin Biol 2008	L. acidophilus LA102,
	L. lactis LA103, S. thermophilus
	LA104
Enck et al., Neurogastroeterol	E. coli DSM 17252,
Motility 2008	<i>Enterococcus faecalis</i> DSM 16440
Andriulli et al., J Clin Gastroenterol 2008	L. paracasei B 21060, L. paracasei B21070,
	L. gasseri B21090
Williams et al., Alim Pharmacol	L. acidophilus CUL60,
Ther 2009	L. acidophilus CUL21,
	B. lactis CUL34, B. bifidum CUL20
Enck et al., Z Gastroenterol 2009	E.coli DSM17252
Kalman <i>et al.</i> , BMC Gastroenterol 2009	Bacillus coagulans GBI-30
Guandalini <i>et al.</i> , J Pediatr Gastroenterol Nutr 2010	VSL#3 preparation
Romano <i>et al.,</i> J Paediatr Child Health 2010	L. reuteri DSM 17938
Francavilla et al., Pediatrics 2010	Lactobacillus GG
Guglielmetti <i>et al.,</i> Aliment Pharmacol Ther 2011	B. bifidum MIMBb75
Guerra <i>et al.</i> , World J Gastroenterol 2011	B. longum (clinical isolate)
Tarrerias et al., Dig Dis 2011	Lactobacillus LB

VSL#3[®]: B. breve, B. longum, B. infantis, L. acidophilus, L.plantarum, L. paracasei, L. bulgaricus, S. thermophilus. visceral perception comes from experiments where microbiota from healthy NIH Swiss mice was deliberately perturbed by administration of non-absorbable antibiotics. This treatment resulted in gut dysbiosis characterized by increased levels of Enterobacteriae and decrease in Lactobacilli and Bacteroides, and lowgrade gut inflammation. Mice also developed increased visceral perception in response to colorectal balloon distension accompanied by up-regulation of SP in the myenteric plexus.⁶⁵ Interestingly, visceral hyperalgesia and enteric SP levels normalized after treatment with Lactobacillus paracasei NCC2461. The same probiotic was shown to also reverse rectal hyperalgesia in maternally deprived rats.⁶⁶ The effect of probiotics on pain mechanisms extends beyond an effect on enteric nerves as Lactobacillus farciminis was shown to ameliorate stress-induced visceral hyperalgesia and affect neural activation patterns assessed by c-fos staining in the sacral spine, paraventricular nucleus of hypothalamus and medial amygdaloid nucleus.67

Several probiotic bacteria have been tested for their ability to decrease pain perception during colorectal distension. Bifidobacterium infantis 35624 was shown to be effective in reducing visceral pain in both visceral normosensitive (Sprague-Dawley) and visceral hypersensitive (Wistar-Kyoto) rats.⁶⁸ Lactobacillus reuteri is a well- characterized strain with respect to its antinociceptive effect: its administration decreased response to colorectal distension in rats⁶⁹ and prevented hyperexcitability of colonic DRG neurons induced by noxious stimuli.⁷⁰ A subsequent study showed that L. reuteri increases excitability and number of action potentials per depolarizing pulse, decreases calciumdependent potassium channel opening and decreases the slow after hyperpolarization in sensory AH neurons.⁷¹ It is important to note that the effect of a given probiotic on visceral pain is likely site specific, as a recent study has shown that L. reuteri but not Lactobacillus plantarum alleviates the response to gastric distension in rats, although both of them have previously been shown to be effective anti-nociceptive agents in colorectal distension.72

The mechanisms of action of specific probiotics on visceral pain perception and ENS modulation are likely to be multiple and strain specific. The D-alanine content of lipoteichoic acid was found to be crucial for *Lactobacillus plantarum*-mediated protection from visceral pain perception in rats.⁷³ Another probiotic, *Lactobacillus acidophilus* NCFM induced the expression of opioid and cannabinoid receptors in intestinal epithelial cells, and modulated analgesic functions in the gut of healthy rats.⁷⁴

Challenges of studying the role of microbiota in the gut-brain axis

Although here is accumulating evidence for bacteria playing an important role in gut-brain axis, our knowledge is limited by our ability to identity the key players in this complex bacterial community. Most of the evidence comes from animal experiments where the microbiota is less diverse than in humans and can be controlled experimentally (e.g. gnotobiotic conditions). In the last decade, significant advancement has been provided by the availabilitity of molecular techniques, such as deep sequencing. These techniques enable us to determine bacteria at the species level. However, the task to identify a microorganism responsible for effects on the CNS is challenging. First, the key bacterium could be underrepresented in the community and could be missed when scanning for total microbiota composition. Also, results are influenced by the technique employed in the analysis of microbiota (454 pyrosequencing, Illumina platform, microarrays). The choice of technique can also make the comparison between individual studies difficult. Second, even if we identify the key microorganism(s) it may be difficult to culture and to determine its characteristics. Last, it is possible that the action of microbiota on the CNS is not related to a specific bacterium or group of bacteria, but rather to their metabolic activity. This metabolic profile may be influenced by environmental factors such as diet. stress or inflammation. Thus, a metabolomic approach would be more likely to yield answers than identification of single bacteria. Despite these methodological set-backs, our understanding of microbial-host interactions is rapidly advancing and this will likely

translate into our ability to diagnose and treat patients with both gut and brain disorders.

CONCLUSIONS

While clinical observation and psychiatric co-morbidity in various chronic intestinal disorders support a role of the intestinal microbiota in gut-brain axis communication, the strongest evidence for a role of microbes as signaling components in the gut-brain axis comes form from animal studies using perturbation of the microbiota by antimicrobials and gnotobiotic models. Mechanisms of communication are likely to be multiple and involve neural, humoral and inflammatory pathways, depending on host and environmental factors. Animal studies will be crucial to continue to provide mechanistic insight and proof-of concept studies. However, there is need for the concepts generated in animal models to be translated to the human in the future.

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REFERENCES

- 1 Banks WA. Evidence for a cholecystokinin gut-brain axis with modulation by bombesin. *Peptides* 1980; 1: 347–51.
- 2 Mayer EA. Gut feelings: the emerging biology of gut-brain communication. *Nat Rev Neurosci* 2011; **12**: 453–66.
- 3 O'Mahony SM, Hyland NP, Dinan TG, Cryan JF. Maternal separation as a model of brain-gut axis dysfunction. *Psychopharmacology* 2011; **214**: 71–88.
- 4 Eckburg PB, Bik EM, Bernstein CN *et al.* Diversity of the human intestinal microbial flora. *Science* 2005; **308**: 1635–8.

- 5 Hooper LV, Macpherson AJ. Immune adaptations that maintain homeostasis with the intestinal microbiota. *Nat Rev Immunol* 2010; **10**: 159–69.
- 6 Bäckhed F, Ley RE, Sonnenburg JL et al. Host-bacterial mutualism in the human intestine. *Science* 2005; **307**: 1915–20.
- 7 Husebye E, Hellstrom PM, Sundler F et al. Influence of microbial species on small intestinal myoelectric activity and transit in germ-free rats. Am J Physiol Gastrointest Liver Physiol 2001; **280**: G368–80.
- 8 Verdu EF, Collins SM. Microbial-gut interactions in health and disease. Irritable bowel syndrome. *Best Pract*

Res Clin Gastroenterol 2004; **18**: 315–21.

- 9 Hooper LV, Gordon JI. Commensal host-bacterial relationships in the gut. *Science* 2001; **292**: 1115–8.
- 10 Dumas ME, Barton RH, Toye A *et al.* Metabolic profiling reveals a contribution of gut microbiota to fatty liver phenotype in resistant mice. *Proc Natl Acad Sci USA* 2006; **103**: 12511–6.
- 11 Zoetendal E, Akkermans AD, DeVos WM. Temperature gradient gel electrophoresis analysis of 16S rRNA from human fecal samples reveals stable and host-specific communities of active bacteria. *Appl Environ Microbiol* 1998; **64**: 3854–9.

- 12 Gronlund MM, Lehtonen OP, Eerola E, Kero P. Fecal microflora in healthy infants born by different methods of delivery: permanent changes in intestinal flora after cesarean delivery. J Pediatr Gastroenterol Nutr 1999; 28: 19–25.
- 13 Martin R, Heilig GH, Zoetendal EG et al. Diversity of the Lactobacillus group in breast milk and vagina of healthy women and potential role in the colonization of the infant gut. J Appl Microbiol 2007; 103: 2638–44.
- 14 Coppa GV, Bruni S, Morelli L *et al.* The first prebiotics in humans: human milk oligosaccharides. *J Clin Gastroenterol* 2004; **38**: S80–3.
- 15 Lewis S, Cochrane S. Alteration of sulfate and hydrogen metabolism in the human colon by changing intestinal transit rate. *Am J Gastroenterol* 2007; **102**: 624–33.
- 16 Tannock GW, Munro K, Harmsen HJM et al. Analysis of the fecal microflora of human subjects consuming a probiotic containing Lactobacillus rhamnosus DR20. Appl Environ Microbiol 2000; 66: 2578–88.
- 17 Zoetendal EG, Akkermans ADL, Vliet WMA *et al.* The Host Genotype Affects the Bacterial Community in the Human Gastronintestinal Tract. *Microb Ecol Health Dis* 2001; **13**: 129–34.
- 18 Arumugam M, Raes J, Pelletier E et al. Enterotypes of the human gut microbiome. Nature 2011; 473: 174–80.
- 19 Wu GD, Chen J, Hoffmann C et al. Linking long-term dietary patterns with gut microbial enterotypes. *Sci*ence 2011; 334: 105–8.
- 20 McCartney AL, Wenzhi W, Tannock GW. Molecular analysis of the composition of the bifidobacterial and lactobacillus microflora of humans. *Appl Environ Microbiol* 1996; 62: 4608–13.
- 21 Bartosch S, Fite A, Macfarlane GT, McMurdo ME. Characterization of bacterial communities in feces from healthy elderly volunteers and hospitalized elderly patients by using realtime PCR and effects of antibiotic treatment on the fecal microbiota. *Appl Environ Microbiol* 2004; 70: 3575–81.
- 22 Kleessen B, Schroedl W, Stueck M et al. Microbial and immunological responses relative to high-altitude exposure in mountaineers. *Med Sci Sports Exerc* 2005; **37**: 1313–8.

- 23 McBurney WT, McCartney A, Apun A et al. Perturbation of the enterobacterial microflora detected by molecular analysis. *Microb Ecol Health Dis* 1999; 11: 75–9.
- 24 Löfmark S, Jernberg C, Jansson JK, Edlund C. Clindamycin-induced enrichment and long-term persistence of resistant *Bacteroides* spp. and resistance genes. *J Antimicrob Chemother* 2006; **58**: 1160–7.
- 25 Jernberg C, Löfmark S, Edlund C, Jansson JK. Long-term ecological impacts of antibiotic administration on the human intestinal microbiota. *ISME J* 2007; **1**: 56–66.
- 26 De FilippoC, Cavalieri D, Di Paola M *et al.* Impact of diet in shaping gut microbiota revealed by a comparative study in children from Europe and rural Africa. *Proc Natl Acad Sci USA* 2010; **107**: 14691–6.
- 27 Maukonen J, Satokari R, Mättö J *et al.* Prevalence and temporal stability of selected clostridial groups in irritable bowel syndrome in relation to predominant faecal bacteria. *J Med Microbiol* 2006; **55**: 625–33.
- 28 Mättö J, Maunuksela L, Kajander K et al. Composition and temporal stability of gastrointestinal microbiota in irritable bowel syndrome – a longitudinal study in IBS and control subjects. FEMS Immunol Med Microbiol 2005; 43: 213–22.
- 29 Martinez C, Antolin M, Santos J et al. Unstable composition of the fecal microbiota in ulcerative colitis during clinical remission. Am J Gastroenterol 2008; 103: 643–8.
- 30 Scanlan PD, Shanahan F, O'Mahony C, Marchesi JR. Culture-independent analyses of temporal variation of the dominant fecal microbiota and targeted bacterial subgroups in Crohn's disease. J Clin Microbiol 2006; 44: 3980–8.
- 31 Bass NM. Review article: the current pharmacological therapies for hepatic encephalopathy. *Aliment Pharmacol Ther* 2007; **25**(Suppl 1): 23–31.
- 32 Parracho HM, Bingham MO, Gibson GR, McCartney AL. Differences between the gut microflora of children with autistic spectrum disorders and that of healthy children. *J Med Microbiol* 2005; **54**: 987–91.
- 33 Sandler RH, Finegold SM, Bolte ER et al. Short-term benefit from oral vancomycin treatment of regressiveonset autism. J Child Neurol 2000; 15: 429–35.

- 34 Posey DJ, Kem DL, Swiezy NB et al. A pilot study of D-cycloserine in subjects with autistic disorder. Am J Psychiatry 2004; 161: 2115–7.
- 35 Mehdi S. Antibiotic-induced psychosis: a link to D-alanine? *Med Hypotheses* 2010; **75**: 676–7.
- 36 Ledochowski M, Sperner-Unterweger B, Fuchs D. Lactose malabsorption is associated with early signs of mental depression in females: a preliminary report. *Dig Dis Sci* 1998; 43: 2513–7.
- 37 Ledochowski M, Widner B, Sperner-Unterweger B *et al.* Carbohydrate malabsorption syndromes and early signs of mental depression in females. *Dig Dis Sci* 2000; **45**: 1255–9.
- 38 Lyte M, Varcoe JJ, Bailey MT. Anxiogenic effect of subclinical bacterial infection in mice in the absence of overt immune activation. *Physiol Behav* 1998; 65: 63–8.
- 39 Gaykema RP, Goehler LE, Lyte M. Brain response to cecal infection with *Campylobacter jejuni*: analysis with Fos immunohistochemistry. *Brain Behav Immun* 2004; **18**: 238– 45.
- 40 Goehler LE, Park SM, Opitz N *et al. Campylobacter jejuni* infection increases anxiety-like behavior in the holeboard: possible anatomical substrates for viscerosensory modulation of exploratory behavior. *Brain Behav Immun* 2008; **22**: 354–66.
- 41 Lyte M, Li W, Opitz N *et al.* Induction of anxiety-like behavior in mice during the initial stages of infection with the agent of murine colonic hyperplasia *Citrobacter rodentium. Physiol Behav* 2006; **89**: 350–7.
- 42 Goehler LE, Gaykema RP, Opitz N et al. Activation in vagal afferents and central autonomic pathways: early responses to intestinal infection with Campylobacter jejuni. Brain Behav Immun 2005; **19**: 334–44.
- 43 Bercík P, De GiorgioR, Blennerhassett P et al. Immune-mediated neural dysfunction in a murine model of chronic Helicobacter pylori infection. *Gastroenterology* 2002; **123**: 1205–15.
- 44 Bercik P, Verdú EF, Foster JA *et al.* Role of gut-brain axis in persistent abnormal feeding behavior in mice following eradication of *Helicobacter pylori* infection. *Am J Physiol Regul Integr Comp Physiol* 2009; **296**: R587–94.
- 45 Sudo N, Chida Y, Aiba Y *et al.* Postnatal microbial colonization programs the hypothalamic-pituitary-adrenal

system for stress response in mice. *J Physiol* 2004; **558**: 263–75.

- 46 Heijtz RD, Wang S, Anuar F et al. Normal gut microbiota modulates brain development and behavior. Proc Natl Acad Sci U S A 2011; 108: 3047– 52.
- 47 Neufeld KM, Kang N, Bienenstock J, Foster JA. Reduced anxiety-like behavior and central neurochemical change in germ-free mice. *Neurogastroenterol Motil* 2011; 23: 255–64.
- 48 Hooper LV, Wong MH, Thelin A *et al.* Molecular analysis of commensal host-microbial relationships in the intestine. *Science* 2001; 291: 881–4.
- 49 Bercik P, Denou E, Collins J et al. The intestinal microbiota affect central levels of brain-derived neurotropic factor and behavior in mice. *Gastroenterology* 2011, **141**: 599–609.
- 50 Conti LH, Costello DG, Martin LA et al. Mouse strain differences in the behavioral effects of corticotropinreleasing factor (CRF) and the CRF antagonist alpha-helical CRF9-41. *Pharmacol Biochem Behav* 1994; 48: 497–503.
- 51 Kalinichev M, Bate ST, Coggon SA *et al.* Locomotor reactivity to a novel environment and sensitivity to MK-801 in five strains of mice. *Behav Pharmacol* 2008; **19**: 71–5.
- 52 Gareau MG, Wine E, Rodrigues DM et al. Bacterial infection causes stress-induced memory dysfunction in mice. *Gut* 2011; **60**: 307–17.
- 53 Li W, Dowd SE, Scurlock B et al. Memory and learning behavior in mice is temporally associated with diet-induced alterations in gut bacteria. *Physiol Behav* 2009; **96**: 557–67.
- 54 Folks DG. The interface of psychiatry and irritable bowel syndrome. *Curr Psychiatry Rep* 2004; **6**: 210–5.
- 55 Whitehead WE, Palsson O, Jones KR. Systematic review of the comorbidity of irritable bowel syndrome with other disorders: what are the causes and implications? *Gastroenterology* 2002; **122**: 1140–56.
- 56 Harter MC, Conway KP, Merikangas KR. Associations between anxiety disorders and physical illness. Eur

Arch Psychiatry Clin Neurosci 2003; 253: 313–20.

- 57 Fuller-Thomson E, Sulman J. Depression and inflammatory bowel disease: findings from two nationally representative Canadian surveys. *Inflamm Bowel Dis* 2006; **12**: 697–707.
- 58 Walker JR, Ediger JP, Graff LA et al. The Manitoba IBD cohort study: a population-based study of the prevalence of lifetime and 12-month anxiety and mood disorders. Am J Gastroenterol 2008; 103: 1989–97.
- 59 Bercik P, Verdu EF, Foster JA *et al.* Chronic gastrointestinal inflammation induces anxiety-like behavior and alters central nervous system biochemistry in mice. *Gastroenterol*ogy 2010; **139**: 2102–12. e1.
- 60 Bercik P, Park AJ, Sinclair D *et al.* The anxiolytic effect of *Bifidobacterium longum* NCC3001 involves vagal pathways for gut-brain communication. *Neurogastroenterol Motil* 2011; **23**: 1132–9.
- 61 Desbonnet L, Garrett L, Clarke G, Bienenstock J, Dinan TG. The probiotic *Bifidobacteria infantis*: an assessment of potential antidepressant properties in the rat. J Psychiatr Res 2008; **43**: 164–74.
- 62 Desbonnet L, Garrett L, Clarke G et al. Effects of the probiotic *Bifido*bacterium infantis in the maternal separation model of depression. Neuroscience 2010; **170**: 1179–88.
- 63 Bravo JA, Forsythe P, Chew MV et al. Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. *Proc Natl Acad Sci U S A* 2011; **108**: 16050–5.
- 64 O'Mahony L, McCarthy J, Kelly P et al. Lactobacillus and Bifidobacterium in irritable bowel syndrome: symptom responses and relationship to cytokine profiles. Gastroenterology 2005; **128**: 541–51.
- 65 Verdú EF, Bercik P, Verma-Gandhu M et al. Specific probiotic therapy attenuates antibiotic induced visceral hypersensitivity in mice. *Gut* 2006; 55: 182–90.

- 66 Eutamene H, Lamine F, Chabo C et al. Synergy between Lactobacillus paracasei and its bacterial products to counteract stress-induced gut permeability and sensitivity increase in rats. J Nutr 2007, 137: 1901–7.
- 67 Ait-Belgnaoui A, Eutamene H, Houdeau E et al. Lactobacillus farciminis treatment attenuates stress-induced overexpression of Fos protein in spinal and supraspinal sites after colorectal distension in rats. Neurogastroenterol Motil 2009; **21**: 567–73.
- 68 McKernan DP, Fitzgerald P, Dinan TG, Cryan JF. The probiotic *Bifidobacterium infantis* 35624 displays visceral antinociceptive effects in the rat. *Neurogastroenterol Motil* 2010; 22: 1029–35.
- 69 Kamiya T, Wang L, Forsythe P *et al.* Inhibitory effects of *Lactobacillus reuteri* on visceral pain induced by colorectal distension in Sprague-Dawley rats. *Gut* 2006; **55**: 191–6.
- 70 Ma X, Mao YK, Wang B et al. Lactobacillus reuteri ingestion prevents hyperexcitability of colonic DRG neurons induced by noxious stimuli. Am J Physiol Gastrointest Liver Physiol 2009; 296: G868–75.
- 71 Kunze WA, Mao YK, Wang B et al. Lactobacillus reuteri enhances excitability of colonic AH neurons by inhibiting calcium-dependent potassium channel opening. J Cell Mol Med 2009; 13: 2261–70.
- 72 Duncker SC, Kamiya T, Wang L et al. Probiotic Lactobacillus reuteri alleviates the response to gastric distension in rats. J Nutr 2011; 141: 1813–8.
- 73 Duncker SC, Wang L, Hols P, Bienenstock J. The D-alanine content of lipoteichoic acid is crucial for *Lactobacillus plantarum*-mediated protection from visceral pain perception in a rat colorectal distension model. *Neurogastroenterol Motil* 2008; 20: 843– 50.
- 74 Rousseaux C, Thuru X, Gelot A et al. Lactobacillus acidophilus modulates intestinal pain and induces opioid and cannabinoid receptors. Nat Med 2007; 13: 35–7.